

An Alternative Reaction Course in O-Glycosidation with O-Glycosyl Trichloroacetimidates as Glycosyl Donors and Lewis Acidic Metal Salts as Catalyst: Acid–Base Catalysis with Gold Chloride-Glycosyl Acceptor Adducts

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Supporting Information

ABSTRACT: Gold(III) chloride as catalyst for *O*-glycosyl trichloroacetimidate activation revealed low affinity to the glycosyl donor but high affinity to the hydroxy group of the acceptor alcohol moiety, thus leading to catalyst–acceptor adduct formation. Charge separation in this adduct, increasing the proton acidity and the oxygen nucleophilicity, permits donor activation and concomitant acceptor transfer in a hydrogen-bond mediated S_N2 -type transition state. Hence, the sequential binding between acceptor and catalyst and then with the glycosyl donor enables self-organization of an



ordered transition-state. This way, with various acceptors, even at temperatures below -60 °C, fast and high yielding glycosidations in high anomeric selectivities were recorded, showing the power of this gold(III) chloride acid–base catalysis. Alternative reaction courses via hydrogen chloride or HAuCl₄ activation or intermediate generation of glycosyl chloride as the real donor could be excluded. With partially *O*-protected acceptors, prone to bidentate ligation to gold(III) chloride, particularly high reactivities and anomeric selectivities were observed. Gold(I) chloride follows the same catalyst–acceptor adduct driven acid–base catalysis reaction course.

INTRODUCTION

For the activation of O-glycosyl trichloroacetimidate glycosyl donors in the presence of glycosyl acceptors (for instance, hydroxy compounds), mainly catalytic amounts of trimethylsilyl trifluoromethanesulfonate (TMSOTf) or borontrifluoride etherate (BF₃·Et₂O), respectively, are employed.¹⁻⁵ There is good evidence that the activation of O-glycosyl trichloroacetimidates is effected by direct attack of the Lewis acid or the Brønsted acid catalyst (Cat) at the imidate nitrogen (Scheme 1). Thus, via an O-glycosyl trichloroacetimidate-catalyst transition state, a glycosyl oxocarbenium ion (short: glycosyl cation) intermediate and trichloroacetamide (TCAA) as leaving group are generated. The glycosyl cation reacts directly with the acceptor to the glycoside; thus, the anomeric selectivity is essentially determined by steric and/or stereoelectronic effects of the glycosyl donor. Alternatively, the evolving glycosyl cation is stabilized by anchimeric assistance, the solvent or by other nucleophiles present in the reaction mixture; this way these effectors influence the yield and particularly the anomeric selectivity in the glycosidation step.

For the *O*-glycosyl trichloroacetimidate activation, also quite a few Lewis acidic metal salts were investigated and the same reaction course was envisaged. Worth mentioning are Ni-(II)⁶⁻¹¹ and Pd(II) salts,^{12–14} ZnBr₂,¹⁵ and MgBr₂·Et₂O in the presence and absence of a Brønsted acid,¹⁶ AuCl,¹⁷ InCl₃,¹⁸ InBr₃,¹⁸ In(OTf)₃,¹⁸ Sm(OTf)₃,¹⁹ Yb(OTf)₃,²⁰ Sn(OTf)₂,²¹ and AgOTf.^{22,23}

In accordance with this general reaction scheme, catalysts with high affinity to the glycosyl donor leaving group (as for instance TMSOTf or BF3·Et2O, respectively) exhibit also disadvantages: As these catalysts generate the highly reactive glycosyl cation intermediate irrespective of the presence or absence of an acceptor, they permit competing reactions that may lead to loss of the glycosyl donor properties. To overcome this problem, recently a variation of this reaction scheme was designed by choosing Lewis acid catalysts (i) with low affinity to the glycosyl donor leaving group, thus not directly generating glycosyl cations, however (ii) with high affinity to the glycosyl acceptor, generating in a rapid and reversible reaction an acceptor-catalyst adduct RO-Cat-H (Scheme 2).²⁴ The ensuing (iii) increase in proton acidity and (iv) increase of acceptor nucleophilicity in this adduct should provide the glycosyl donor activation via proton transfer of the acidified acceptor proton to the glycosyl donor and concomitantly to acceptor transfer from the negatively charged $[RO-Cat]^-$ moiety to the incipient glycosyl cation in an S_N2type transition state. Thus, in an hydrogen-bond mediated selforganizing intramolecular acid-base catalyzed reaction, the glycoside should be generated in high yield and generally high anomeric selectivity.^{24,25} This reaction design could be verified

Received: July 28, 2015 Published: September 11, 2015 Scheme 1. Glycosyl Cation Generation and Eventual Concomitant Intra/Intermolecular Stabilization and Final Transformation into Glycoside



Scheme 2. Acceptor-Catalyst Adduct Formation Leads to Acid-Base Catalyzed Glycosidation with a Catalyst Having (i) Low Affinity to the Donor but (ii) High Affinity to the Acceptor



by us, for instance, for $PhBF_2$, Ph_2BF , or $PhSiF_3$, as catalysts.^{24,26–28}

As some of the metal salts employed for O-glycosyl trichloroacetimidate activation are relatively weak Lewis acids, the generally claimed direct activation of the glycosyl donor by these catalysts is questionable. Rather a high affinity to the acceptor hydroxy group is expected, that favors the pathway outlined in Scheme 2. In order to study this question, that has a major impact on product formation, gold(III) chloride, with its strong tendency to form dimers (2 $AuCl_3 \rightleftharpoons Au_2Cl_6$) and in addition to position four ligands in a square-planar orientation,²⁹ seemed to be an ideal catalyst candidate: Dedimerization of Au₂Cl₆ by alcohols (HOR) should lead to an RO^{δ} -AuCl₃-H^{δ +} adduct with strong charge separation for proton-induced O-glycosyl trichloroacetimidate activation and sufficient propensity to concomitant alkoxide transfer to the incipient glycosyl cation (Scheme 2). Binding of the acceptor in a square-planar orientation will lead to a less hindered transition state than in a tetrahedral orientation of the four ligands around the central atom (as for instance in the PhF₂B· HOR or the Ph₂FB·HOR adducts).

Gold(III) chloride mediated activation of glycosyl donors having alkyne moiety containing leaving groups has been reported.^{30–35} Very recently, Vankar et al. also reported on the activation of *O*-glycosyl trichloroacetimidates by gold(III) chloride in the presence of phenylacetylene.³⁶ Some annotations on their results will be discussed below.

RESULTS AND DISCUSSION

Reaction Course of Gold(III) Chloride Catalysis. In order to cope with this task, first the interaction of gold(III) chloride with glycosyl donor 1α (Scheme 3) in CDCl₃ as solvent was studied by NMR spectroscopy: Interaction of 1α with 0.1 equiv of gold(III) chloride at -50 °C was, if at all, only

Scheme 3. Potential Alternative Reaction Pathways. ($R = CHMe_2$)



very weak, as indicated by NMR shift differences (Figure 1); 1α was also not decomposed (Table 1, entry 1). Only by raising



Figure 1. ¹H NMR of donor 1α and a mixture of donor 1α and gold(III) chloride (0.1 equiv) in CDCl₃ at -50 °C.

the temperature (entry 2) or by adding 1.0 equiv of gold(III) chloride (entry 3) slow decomposition of 1α took place. However, interaction of gold(III) chloride with an acceptor, as for instance isopropanol (**A**), was very strong (Figure 2), thus demonstrating the formation of the RO–AuCl₃–H adduct (entry 4). Addition of 1α to this mixture at -60 °C, (hence, employing the inverse procedure³⁷ for glycosidation) led practically to exclusive formation of the β -glucoside $2A\beta$ (Table 1, entry 5). Essentially, the same result was obtained with the standard glycosidation procedure, where the donor 1α and the acceptor **A** were dissolved first in dichloromethane (DCM) and then 0.1 equiv of gold(III) chloride was added to the reaction mixture at -70 °C (entry 6). However, addition of



Figure 2. ¹H NMR of isopropanol and a mixture of isopropanol and gold(III) chloride in CDCl₃ at room temperature.

the catalyst to the donor $\mathbf{1\alpha}$ at -70 °C and thereafter adding the acceptor **A** led only to a very sluggish reaction (entry 7) that could be accelerated by raising the temperature; yet, this resulted in partial decomposition of $\mathbf{1\alpha}$. Presumably, addition of the catalyst to the donor leads to formation of clusters [AuCl₃·($\mathbf{1\alpha}$)_n, n = 1, 2, 3...] that can only slowly be penetrated by the acceptor molecules, this way slowing down the reaction rate. Cluster formation between catalyst and acceptor has been previously observed in the "inverse procedure";³⁷ however, in this case, donor decomposition is obviously no problem and the mobility of the cluster constituents seems to be high, and therefore, there is only a small effect on the reaction rate.³⁸

Besides the proposed gold(III) chloride mediated acid–base catalysis of this reaction (Scheme 2), alternative routes for product formation are available that had to be investigated (Scheme 3). (a) Reaction of 1α and A under gold(III) chloride catalysis leads only to minor, if any, glycosyl chloride $3\alpha_{,\beta}$ formation (Table 1, entries 5–7). Yet, as an alternative to gold(III) catalysis, acid catalysis of this reaction by hydrogen chloride (HCl), eventually generated from the reaction of

Table 1. Interaction and Reactions between	Glycosyl Donor 1α , Acceptor	A and Gold(III) Chloride as	Catalyst ^a
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	$\begin{array}{c} & OBn \\ BnO \\ BnO \\ BnO \\ A \\ 1\alpha \\ CCl_3 \\ A \end{array} + HO \underbrace{Cat} \\ A \\ \end{array}$	BnO BnO 2A α,β	wO	
entry	reaction procedure	T (°C)	time	result
1	1α (1 equiv) + AuCl ₃ (0.1 equiv)	-50	30 min	no reaction, no decomp
2	1α (1 equiv) + AuCl ₃ (0.1 equiv)	rt	30 min	decomp
3	1α (1 equiv) + AuCl ₃ (1 equiv)	-50	30 min	decomp
4	$A (1 \text{ equiv}) + AuCl_3 (1 \text{ equiv})$	rt	30 min	adduct formation
5	A (1.5 equiv) + AuCl ₃ (0.1 equiv), then 1α (1 equiv)	-70	10 min	2Αβ (90%, $β$)
6	1α (1 equiv) + A (1.5 equiv), then AuCl ₃ (0.1 equiv)	-70	10 min	2Αβ (87%, $β$)
7	1α (1 equiv) + AuCl ₃ (0.1 equiv), then A (1.5 equiv)	-70 to rt	overnight	$2A\alpha/\beta$ (40%, 1:2)
8	A (1.5 equiv) + HCl (0.1 equiv) in Et_2O , then 1α (1 equiv)	-70 to rt	overnight	trace
9	A (1.5 equiv) + HCl (1 equiv) in Et_2O , then 1α (1 equiv)	-70	10 min	2Alpha/eta (20%, 1:3), $3lpha$ (15%)
10	A (1.5 equiv) + HAuCl ₄ (0.1 equiv), then 1α (1 equiv)	-70	10 min	2Aβ (85%), 3α (3%)
11	A (1.5 equiv) + AuCl ₃ (0.1 equiv), then 3α (1 equiv)	-70	3 h	no reaction
12	D (1.5 equiv) + AuCl ₃ (0.1 equiv) in MeCN (0.2 mL), then 1α (1 equiv)	-60	30 min	2Dβ (85%, β)
13	PhCCH (0.1) + AuCl ₃ (0.1 equiv) + D (1 equiv), then 1α (1 equiv)	-70 to rt	overnight	no reaction
14	PhCCH (0.5 equiv) + AuCl ₃ (0.1 equiv) + D (1 equiv), then 1α (1 equiv)	rt	10 min	no reaction
15	1α (1 equiv) + D (1 equiv), then PhCCH (0.5 equiv) + AuCl ₃ (0.1 equiv)	rt	10 min	$2D\alpha/\beta$ (70%, 1:1)

^aEntries 1–4 were carried out in $CDCl_3$ (in the NMR spectrometer); entries 5–15 were carried out in DCM.

gold(III) chloride with the acceptor,³⁹ had to be investigated. Therefore, 0.1 equiv of HCl (Et₂O adduct) were added to a mixture of donor 1α and acceptor A; however, only a trace of a $2A\alpha_{\mu}\beta$ -mixture and some glycosyl chloride 3α were formed (Table 1, entry 8). Thus, it could be concluded that HCl is not a decisive catalyst for the observed fast β -glycoside formation. This was also supported by the fact that higher amounts of HCl led to formation of a product mixture of $2A\alpha\beta$ and 3α (entry 9). (b) Alternatively, the HCl-addition product of gold(III) chloride that is 0.1 equiv of HAuCl₄, was investigated as catalyst (entry 10). As expected, beside glycoside $2A\beta$ some glycosyl chloride 3α was now obtained. Hence, after consumption of some HCl from HAuCl₄ subsequently the above-discussed intramolecular gold(III) chloride mediated acid-base catalysis is effective in this reaction. Yet, these studies also exhibit that any compound in the reaction mixture with affinity to gold(III) chloride will inhibit the glycosidation, as found for substrates and/or reagents containing impurities. (c) Finally, the question remains: is glycosyl chloride $3\alpha_{\mu}\beta$, generated as intermediate and then attacked by gold(III) chloride providing the glycosyl cation and stable tetrachloroaurate (AuCl₄⁻), the real glycosyl donor? To this end, glycosyl chloride 3α was activated by 0.1 equiv of gold(III) chloride in the presence of acceptor A; however, no reaction was observed (entry 11). This result further supports the proposed acid-base catalysis of the glycosidation reaction between 1α and A by gold(III) chloride.

As the gold(III) chloride catalyzed reaction is sensitive to the presence of impurities that are capable of binding competitively to gold(III), it was of interest to study the influence of solvents with ligand properties. To this end, the influence of acetonitrile was studied. As expected, this molecule lowered the glycosidation rate, however not the final glycosidation result (entry 12).

In this context, also the influence of phenylacetylene was studied³⁶ because of the high alkynophilicity of gold(III). Application of the inverse glycosidation procedure at -70 °C in the presence of phenylacetylene led to total inhibition of the glycosidation reaction; this was even so at room temperature (entries 13, 14; D was selected as acceptor). However, the reaction proceeded nicely with the standard glycosidation procedure;³⁶ that is, first the donor, acceptor, and phenylacetylene were dissolved at room temperature, and then gold(III) chloride was added (entry 15). Therefore, NMR studies were performed at room temperature, that showed between gold(III) chloride and phenylacetylene complex formation. Following the inverse procedure, addition of isopropanol to this mixture leads to a new complex that can also be obtained by adding first isopropanol and then phenylacetylene to gold(III) chloride (see the Supporting Information (SI)). However, this new complex is unable to activate O-glycosyl trichloroacetimidate glycosyl donor 1α (Table 1, entries 13, 14). Hence, it seems that, under the standard glycosidation procedure, formation of this inactive catalyst-phenylacetylene-acceptor complex is slower than the direct activation of glycosyl donor 1α by the catalyst. (Table 1, entry 15).

Applications to Glycoside Synthesis. The application of this gold(III) chloride acid–base catalysis to various acceptors (Table 2, A–H) having one unprotected hydroxy group and 1α as donor led to excellent glycosidation yields and generally to high β -selectivities, particularly with reactive acceptors (Table 2, 2A–H). Worth mentioning are the glycosidation results obtained with 2-*O*- and 4-*O*-unprotected acceptors E and G,



	PC		AuCl ₃ 4Å MS	2
		donor CCl ₃	anhydrous DCM	→~ OR) luct
entry	donor	acceptor	product	yield β/α ratio ^e
1 ^{a,b}	1α)—oh A	BnO BnO BnO BnO 2Aβ	90% β
2 ^a	1α	=∕_OH B	BnO BnO BnO BnO BnO BnO BnO BnO	88% β
3ª	1α	с он		91% β
4 ^c	1α	HO BnO BnO BnO D OMe	BNO BNO BNO BNO BNO BNO BNO BNO BNO BNO	84% β:α= 12:1
5 ^{c,d}	F 1α	BnO E HO OMe	BnO	87% β:α= 7:1
6 ^{c,d}	Γ 1α	HO HO HO OMe	BnO Ph + O O O O O O O O O O O O O O O O O O	93% β:α =4 :1
7 ^{c,d}	1α	BnO HO BnO BnO G OMe	$\begin{array}{c} BnO\\ BnO\\ BnO\\ BnO\\ BnO\\ BnO\\ BnO\\ 2G\alpha/\beta BnO OMe \end{array}$	80% β:α=6:1
8 ^{c,d}	1α	BnO OBn HO BnO OMe H	$\begin{array}{c} BnO \\ 2H\alpha/\beta^{BnO} \\ OMe \\ 0 \\ Mall \\ BnO \\ 0 \\ Mall \\ Mal$	82% β:α=11:1
9 ^a	4α	A		87% β
10 ^a	4 α	В	BnO OBn BnO 5B β	90% β
11 ^a	4 β	A	BnO BnO BnO	82% β:α=1:15
12 ^a	6α	Α		81% α
BnO BnO Bnt	ο BnO 1α	BnO OBn BnO BnO OBn CCl ₃ BnO O 4α	$\begin{array}{c c} BnO & OBn & BrO \\ BnO & O & NH & BnO \\ CCI_3 & 4\beta & CCI_3 \end{array}$	

^{*a*}Donor/acceptor ratio 1:1.5. ^{*b*}Under strictly anhydrous conditions, the same result was obtained without addition of molecular sieves. ^{*c*}Donor/acceptor ratio 1.5:1. ^{*d*}The reactions were performed at -60 °C. ^{*e*} β/α ratio was obtained from the ¹H NMR spectra.

respectively, exhibiting the high reactivity of these hydroxy groups that for steric reasons were found less reactive (particularly the 4-hydroxy group) in the PhBF₂ or Ph₂BF catalyzed reactions²⁴ (Scheme 2).

Similar results were obtained with 2,3,4,6-tetra-O-benzyl- α -D-galactopyranosyl trichloroacetimidate 4α as glycosyl donor. With acceptor **A** and **B**, the β -glycosides $5A\beta$ and $5B\beta$, respectively, were generated (Table 2, entries 9, 10). In accordance with the general reaction scheme the corresponding β -D-galactopyranosyl trichloroacetimidate 4β afforded with acceptor **A** mainly the α -glycoside $5A\alpha$ (entry 11). However, neighboring group participation, as in 2-O-benzoyl protected α -D-mannopyranosyl trichloroacetimidate 6α as glycosyl donor,

overrides the formation of the S_N 2-type arrangement between donor and catalyst–acceptor adduct and leads to exclusive formation of the 1,2-*trans* product, that is, the α -glycoside 7A α (entry 12).

In total agreement with the proposed reaction scheme, partly protected acceptors, that act as bidentate ligands, were more reactive and led to higher yields, and even higher β -selectivities than mono-*O*-unprotected acceptors in these gold(III) chloride catalyzed glycosidation reactions (Table 3, reactions with

 Table 3. Gold(III) Chloride Catalyzed Glycosidation with

 Partly O-Protected Acceptors



"Entries 1–6 were carried out with 1.0 equiv of donor, 1.5 equiv of acceptor, and 0.15 equiv of AuCl₃. ^bSmall amounts (<8%) of β -(1–4) linked disaccharide $2\mathbf{I}\beta'$, $2\mathbf{J}\beta'$, $2\mathbf{K}\beta'$, $2\mathbf{L}\beta'$, and $2\mathbf{N}\beta'$ were also detected. ^cTrisaccharide $2\mathbf{M}\beta'$ was obtained in 16% yield.

acceptors I–N). Obviously, this is due to binding of the acceptor via two oxygens to gold(III) under release of one molecule of HCl. However, as only 0.1 equiv of gold(III) chloride were added, the amount of released HCl had practically no effect on the reaction result (see Table 1, entry 8). Thus, from 4,6-O-unprotected glucopyranoside I the β -(1–6)-linked disaccharide **2I** β and from 3,4-O-unprotected α -D-galactopyranoside J, the β -(1–3)-linked disaccharide **2J** β was practically exclusively obtained (entries 1, 2).



Surprisingly, with the closely related β -D-galactopyranoside K as acceptor besides the β -(1-3)-linked disaccharide $2K\beta$ as main product, also the β -(1-4)-linked disaccharide $2K\beta'$ was found as minor product (entry 3). Though acceptors K and L are pseudoenantiomeric, similar regio- and stereoselectivity was observed for acceptor L leading to $2L\beta$ and $2L\beta'$ (entry 4). Hence, there is, if at all, only a minor effect of the acceptor stereochemistry on the glycosidation result. The 2,3,4-O-

unprotected acceptor **M** afforded, as expected, the β -(1-3)-linked disaccharide **2M\beta** as main product. However, as minor product, β -(1-2)-/ β -(1-3)-linked trisaccharide **2M\beta**' was also obtained (entry 5). Linkage to the 4-position was not observed, as it is disfavored due to the presence of the bulky *tert*-butyldiphenylsilyl (TBDPS) 6-*O* protecting group, thus exhibiting how to control the regioselectivity in these reactions. This was also found for the reaction of acceptor **N** where mainly **2N\beta** was obtained (entry 6).

The glycosidation rate difference between mono *O*unprotected and di-*O*-unprotected bidentate ligands was established through a competition experiment between structurally closely related acceptors H and K under the general glycosidation conditions (see Tables 2 and 3). As expected, with excess H and K (1.5 equiv each) from the total disaccharide yield of 81%, a 1:6 ratio of $2H\beta$ vs $2K\beta + 2K\beta'$ was obtained. Hence, the initial glycosidation rate difference is about 1:10 in favor of diol K (Scheme 4). Contrary to the

Scheme 4. Competitive Reaction of 1α between Acceptor K and H Catalyzed by Gold(III) Chloride



results with gold(III) chloride, use of PhBF₂, Ph₂BF, and PhSiF₃ as acid–base catalysts led to a decrease in reaction rate and yield with di-O-unprotected bidentate ligands.²⁴ Thus, further support is provided that gold(III) chloride is available to bidentate ligation and to lower steric constraints in the glycosidation transition state in a square-planar ligand orientation of the acceptor oxygen(s).

Borinic acid derivatives, accessible to bidentate ligation with the acceptors, have been studied with glycosyl halides as glycosyl donors and their activation by equivalent amounts of silver oxide.^{40,41} However, as the glycosidation procedure is noncatalytic, the high yielding anomeric selectivities are essentially based on neighboring group participation and high regioselectivities are only obtained with the help of bulky protecting groups, the results are not really comparable with the present work, that employs powerful gold(III) chloride acid– base catalysis for the concomitant activation of glycosyl donor and acceptor without resorting to archimeric assistance for anomeric selectivity control.

Gold(I) Chloride as Catalyst. The results obtained with gold(III) chloride raise the question: do other Lewis acidic metal salts employed for *O*-glycosyl trichloroacetimidate activation follow the same reaction course? Due to the close relationship with gold(III) chloride, we looked briefly into the behavior of gold(I) chloride, for which good glycosidation results with varying anomeric selectivities were obtained at room temperature.¹⁷ We noticed that even 1.0 equiv of gold(I) chloride is unable to directly activate and this way decompose *O*-glycosyl trichloroacetimidate 1 α . However, with acceptor **A**, complex formation was observed; following addition of donor 1 α furnished glycoside 2A β even at -60 °C in very good yield (Table 4, entry 1). 3-O-Unprotected acceptor F afforded under

Table 4. Gold(I) Chloride Catalyzed Glycosidations



^{*a*}Donor/acceptor ratio 1:1.5. ^{*b*}Donor/acceptor ratio 1.5:1. ^{*c*} β/α ratio was obtained from the ¹H NMR spectra.

these conditions disaccharide $2F\alpha_{,\beta}$ in 78% yield in a 1:3 ratio (entry 2); this way, the reaction rate difference became obvious, with gold(III) chloride being the more active catalyst. Yet, gold(I) chloride acts also as diol accepting entity, as it led with acceptor K practically to the same result, as obtained with gold(III) chloride (Tables 3 and 4, compare entries 3). This result is either due to expansion of the linear two-coordinate gold(I) complexation to a three-coordinate, trigonal-planar complexation,⁴² or alternatively due to disproportionation of gold(I) chloride to gold(0) and gold(III) chloride.⁴³

CONCLUSION

Lewis acids with low affinity to the glycosyl donor leaving group but with high affinity to the glycosyl acceptor are expected to form catalyst-acceptor adducts that, appropriately selected, permit glycosyl donor activation and concomitant acceptor transfer to the incipient glycosyl cation in an S_N2-type intramolecular fashion. This novel conceptual approach to glycosidations that is facilitated by glycosyl donors accessible to activation by catalysis, as the highly reactive O-glycosyl trichloroacetimidate donors,^{1,3} works particularly well with gold(III) chloride as catalyst. This Lewis acid turned out to be a powerful catalyst by binding first to the alcohol generating an adduct that reacts with the donor to the glycoside via acid-base catalysis, that is, via intramolecular dual catalysis. The superior reactivity of this system even at low temperatures exhibits the power of this approach and permits the use of a broad acceptor range, resulting in high glycoside yields and anomeric selectivities. The access of gold(III) chloride to bidentate ligation of diols and the square-planar arrangement of the ligands turns out to be a further advantage of this catalyst system that seems to hold many more surprises, as the studies with competing ligands for gold(III) and the extension of this catalysis to gold(I) chloride display.

EXPERIMENTAL SECTION

General Procedure for Gold(III) Catalyzed Glycosidation. To a solution of gold(III) chloride (4.4 μ mol) and 4 Å molecular sieves (200 mg) in 2 mL of anhydrous DCM was added acceptor A (66 μ mol) at room temperature. (Note: When substrates, reagents, and solvents are particularly carefully dried, molecular sieves are not required in order to obtain the same results (see Table 2, entry 1)). After cooling down the reaction mixture to -70 °C, highly pure donor 1α (44 μ mol), dissolved in 1 mL of anhydrous DCM, was slowly added into the reaction. The reaction was further stirred for 30 min at this temperature. After the TLC analysis showed the completion of the reaction, the reaction was quenched by addition of triethylamine and diluted with 50 mL of DCM. Then the precipitate was filtered off through a pad of Celite. The organic layer was washed with NaHCO₃ (aq.) (10 mL) and brine (10 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by column chromatography (hexane/acetone or hexane/ethyl acetate) on silica gel.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/jacs.5b07895.

Full experimental details and ¹H and ¹³NMR spectra for all new compounds (PDF)

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Notes

The authors declare no competing financial interest.

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REFERENCES

(1) Schmidt, R. R. Angew. Chem., Int. Ed. Engl. 1986, 25, 212-235.

(2) Handbook of Chemical Glycosylation: Advances in Stereoselectivity and Therapeutic Relevance; Demchenko, A. V., Ed.; Wiley-VCH: Weinheim, 2008.

- (3) Zhu, X.; Schmidt, R. R. Angew. Chem., Int. Ed. 2009, 48, 1900–1934.
- (4) Mydock, L. K.; Demchenko, A. V. Org. Biomol. Chem. 2010, 8, 497-510.

(5) Nigudkar, S. S.; Demchenko, A. V. Chem. Sci. 2015, 6, 2687–2704.

(6) Mensah, E. A.; Nguyen, H. M. J. Am. Chem. Soc. 2009, 131, 8778-8780.

(7) Mensah, E. A.; Yu, F.; Nguyen, H. M. J. Am. Chem. Soc. 2010, 132, 14288–14302.

- (8) Yu, F.; Nguyen, H. M. J. Org. Chem. 2012, 77, 7330-7343.
- (9) Yu, F.; McConnell, M. S.; Nguyen, H. M. Org. Lett. 2015, 17, 2018–2021.
- (10) McConnell, M. S.; Yu, F.; Nguyen, H. M. Chem. Commun. 2013, 49, 4313–4315.
- (11) McConnell, M. S.; Mensah, E. A.; Nguyen, H. M. Carbohydr. Res. 2013, 381, 146–152.
- (12) Yang, J.; Cooper-Vanosdell, C.; Mensah, E. A.; Nguyen, H. M. J. Org. Chem. 2008, 73, 794–800.
- (13) McKay, M. J.; Naab, B. D.; Mercer, G. J.; Nguyen, H. M. J. Org. Chem. 2009, 74, 4705–4711.
- (14) Mensah, E. A.; Azzarelli, J. M.; Nguyen, H. M. J. Org. Chem. 2009, 74, 1650–1657.
- (15) Urban, F. J.; Moore, B. S.; Breitenbach, R. Tetrahedron Lett. 1990, 31, 4421–4424.
- (16) Gould, N. D.; Liana Allen, C.; Nam, B. C.; Schepartz, A.; Miller, S. J. *Carbohydr. Res.* **2013**, *382*, 36–42.
- (17) Goetze, S.; Fitzner, R.; Kunz, H. Synlett **2009**, 2009, 3346–3348.
- (18) Mattson, A. L.; Michel, A. K.; Cloninger, M. J. Carbohydr. Res. 2012, 347, 142–146.

(19) Adinolfi, M.; Barone, G.; Guariniello, L.; Iadonisi, A. *Tetrahedron Lett.* **2000**, *41*, 9005–9008.

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(20) Adinolfi, M.; Barone, G.; Iadonisi, A.; Mangoni, L.; Schiattarella, M. *Tetrahedron Lett.* **2001**, *42*, 5967–5969.

(21) Bartek, J.; Müller, R.; Kosma, P. Carbohydr. Res. 1998, 308, 259–273.

- (22) Douglas, S. P.; Whitfield, D. M.; Krepinsky, J. J. J. Carbohydr. Chem. 1993, 12, 131-136.
- (23) Wei, G.; Gu, G.; Du, Y. J. Carbohydr. Chem. 2003, 22, 385–393.
 (24) Kumar, A.; Kumar, V.; Dere, R. T.; Schmidt, R. R. Org. Lett. 2011, 13, 3612–3615.

(25) Geng, Y.; Kumar, A.; Faidallah, H. M.; Albar, H. A.; Mhkalid, I. A.; Schmidt, R. R. Angew. Chem., Int. Ed. 2013, 52, 10089-10092.

(26) Kumar, A.; Geng, Y.; Schmidt, R. R. Adv. Synth. Catal. 2012, 354, 1489-1499.

(27) Kumar, A.; Schmidt, R. R. Eur. J. Org. Chem. 2012, 2012, 2715–2719.

(28) Kumar, A.; Schmidt, R. R. In *Glycoscience: Biology and Medicine*; Taniguchi, N., Endo, T., Hart, G. W., Seeberger, P. H., Wong, C.-H., Eds.; Springer: Japan, 2014; p 295–303.

(29) Clark, E. S.; Templeton, D. H.; MacGillavry, C. H. Acta Crystallogr. 1958, 11, 284–288.

(30) Rao, B. V.; Manmode, S.; Hotha, S. J. Org. Chem. 2015, 80, 1499–1505.

(31) Hotha, S.; Kashyap, S. J. Am. Chem. Soc. 2006, 128, 9620–9621.
(32) Li, Y.; Yang, X.; Liu, Y.; Zhu, C.; Yang, Y.; Yu, B. Chem. - Eur. J.
2010, 16, 1871–1882.

(33) Tang, Y.; Li, J.; Zhu, Y.; Li, Y.; Yu, B. J. Am. Chem. Soc. 2013, 135, 18396–18405.

(34) Yu, B.; Sun, J.; Yang, X. Acc. Chem. Res. 2012, 45, 1227-1236.

(35) Adhikari, S.; Baryal, K. N.; Zhu, D.; Li, X.; Zhu, J. ACS Catal. **2013**, 3, 57–60.

- (36) Roy, R.; Palanivel, A. K.; Mallick, A.; Vankar, Y. D. Eur. J. Org. Chem. 2015, 2015, 4000-4005.
- (37) Schmidt, R. R.; Toepfer, A. Tetrahedron Lett. 1991, 32, 3353–3356.

(38) The importance of the inhomogeneity of solutions on glycosidation results has been recently also discussed by: Kononov, L. O. *RSC Adv.* **2015**, *5*, 46718–46734.

(39) Wabnitz, T. C.; Yu, J.-Q.; Spencer, J. B. Chem. - Eur. J. 2004, 10, 484-493.

(40) Oshima, K.; Aoyama, Y. J. Am. Chem. Soc. **1999**, 121, 2315–2316.

(41) Gouliaras, C.; Lee, D.; Chan, L.; Taylor, M. S. J. Am. Chem. Soc. **2011**, 133, 13926–13929.

(42) Gimeno, M. C.; Laguna, A. Chem. Rev. 1997, 97, 511-522.

(43) *Modern Gold Catalyzed Synthesis*; Hashmi, A. S. K., Toste, F. D., Eds.; Wiley-VCH: Weinheim, 2012.